



# INFLUENCE OF ACETATE AS MAIN CARBON SOURCE FOR LIPID PRODUCTION VIA OLEAGINOUS YEAST FERMENTATION

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# <u>Concept</u>

Microbial oils are proposed as a suitable alternative to petroleum-based chemistry in terms of environmental preservation. These oils have traditionally been studied using sugar-based feedstock, which implies high costs, substrate limitation, and high contamination risks. In this sense, low-cost carbon sources such as acetate are envisaged as promising building blocks for lipid biosynthesis to produce oil-based bioproducts such as chemicals and drop-in biofuels. Here, we investigated the use of acetate as main carbon source for the production of triacylglycerides (TAGs) using the oleaginous yeast *Yarrowia lipolytica*. In order to enhance TAGs production a genetic strategy was used to obtain recombinant obese strains of *Y. lipolytica* capable of accumulating a high amount of lipids. Batch fermentations and continuous fermentations with cell recycle fed with a diluted acetate solution, were carried out at lab scale using 1.5 L bioreactors. Fermentation conditions were studied at laboratory level to determine the most important parameters influencing lipid accumulation (pH, DO, C/N ratio) as well as alternative carbon sources to increase yield of lipid production.

# **Objectives**

The aim was to evaluate feed rates, growth rates and conversion rates (g/g), titers (g/L), volumetric productivity (g/L/h) and fatty acid composition of produced TAGs using different carbon sources (acetate, acetate+glycerol, glucose).

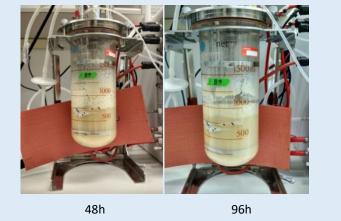
In addition, the performance of the *Y. lipolytica* wild-type (WT) strain and the *Y. lipolytica* modified strain (DGA) have also been compared.

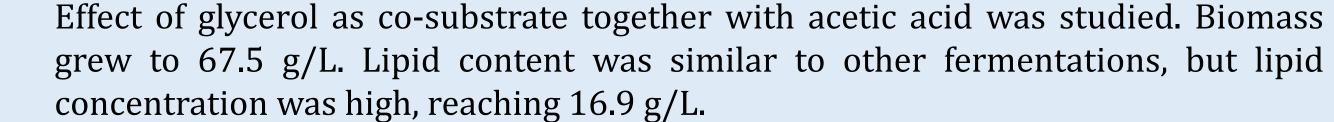
#### **Batch fermentations with acetic acid**

A preliminar design of experiments was carried out to study the influence of three parameters: initial acetate concentration, DO and pH. For each parameter, two reference values were selected. Fermentations were performed during 96 hours.

		рН	DO (%)	Acetate (g/L)	CDW (g/L)	TAGs (g/L)	TAGs/CDW (%)	qTAGs (g/L/h)		4	8h		96h	C16:0 ME
F	F1, WT	5.5	20	15	7.93	1.418	19.85	0.014	50 40			Ŧ	I	C16:0 ME C16:1 ME C18:0 ME C18:1 ME
F	F2, WT	5.5	20	50	8.53	1.358	16.29	0.014	<b>146</b>	I I		-		C18:2 ME
F	F3, WT	8	20	15	13	3.04	12.79	0.031	00 <b>1</b> 0	Ŧ	I	Ŧ	I	
ł	F4, WT	8	20	50	17.4	3.479	19.45	0.036	× 10	24	<b>1</b>	72	96	-
F	F5, WT	8	0	15	1.198	0.1	8.55	0.0011				(hours)		F4 <i>,</i> WT

# <u>Results</u>





Using glucose as sole carbon source, resulting biomass concentration was similar to that obtained with acetic acid. The percentage of TAGs was higher than that obtained so far, reaching 30.8 %.

**Continuous fermentations with other carbon sources** 

TAGs profile distributions were similar to other fermentations.

C16:1 MF C16:0 MF C18:2 MF C18:1 MF C18:0 M

#### **Continuous fermentations with acetic acid**

The operation was based on an initial fed-batch stage and a continuous stage. Initial culture volume was 0.5 L. Substrate was fed until 1.5 L was reached. At this point, continuous operation was started, maintaining a constant volume through continuous extraction of cell-free broth stream (tangential filtration with hollow fiber membrane). Fermentations were performed during 6 days at pH 7 and DO 30%.

	Feed 1	Feed 2	Feed (mL/min)	Biomass (g/L)	% TAGs	TAGs (g/L)	TAGs yield (g/g), Prod (g/L/h)
F5, WT	3 % AA 5 g/L AmSulf C/N=13	3 % AA 0.65 g/L AmSulf C/N=102	0.3	7.9	29.1	2.3	0.20
F6, DGA	3 % AA 5 g/L AmSulf C/N=13	3 % AA 0.65 g/L AmSulf C/N=102	0.25	23.0	22.6	5.2	0.14, 0.04
F7, DGA	3 % AA 5 g/L AmSulf C/N=13	3 % AA 0.65 g/L AmSulf C/N=102	0.88-1.6	9.8	25.2	2.5	0.06, 0.03



F6, DGA

	Feed 1	Feed 2	Feed (mL/min)	DO (%)	рН	Biomass (g/L)	TAGs (%)	TAGs (g/L)	TAGs yield (g/g), Prod (g/L/h)
F8, DGA	3 % AA 3 % glycerol 5 g/L AmSulf C/N=26	3 % AA 3 % glycerol 0.65 g/L AmSulf C/N=204	0.4-0.6	30	7	67.5	25.1	16.9	0.28, 0.10
F9, DGA	6 % AA 6 % glycerol 18 g/L AmSulf C/N=14	6 % AA 6 % glycerol 2.6 g/L AmSulf C/N=100	0.15	30→5	7 to 3 (fast)	26→16.2	12.0	1.9	0.04, 0.03
F10, DGA	3 % AA 3 % glycerol 0.65 g/L AmSulf C/N=204	3 % AA 3 % glycerol C/N = inf	0.09-0.12	30→5	7 to 2.5 (slow)	27.8	24.1	6.7	0.27, 0.06
F11, DGA	(100 g/L glucose in medium)	6 % NaAc 6 % glycerol	0.1	30→10	7	90.5	21.9	19.8	0.177, 0.261
F12, DGA	60 g/L glucose 10 g/L AmSulf C/N=13	60 g/L glucose 1.3 g/L AmSulf C/N=102	0.2	30→5	7 to 2.5 (slow)	22.9	30.8	7	0.11, 0.08
	70       60       50       60       60       50       60       50 <td< th=""></td<>								
		5 20 % 10 0			<sup>tt</sup> 20 % 10 0				

### **Conclusions**

- The use of a continuous fermentation mode with cell recycling using hollow fiber membranes to recirculate the cells while removing exhausted culture medium resulted in increased lipid production.
- Acetate fermentations with modified *Y. lipolytica* strain resulted in a TAGs content of 25% but with low biomass (20-25 g/L) and lipid titer (6.2 g/L).
- Acetate fermentations with modified Y. lipolytica strain using glycerol as a co-substrate resulted in similar TAGs accumulation (%) but higher biomass (67.5 g/L) and therefore
  higher lipid titer (16.9 g/L).
- Glucose fermentation does not provide more TAGs if the nitrogen concentration is not low enough.
- Glucose fermentation with stress conditions such as low DO (5%) and low pH (2.8) during the 2<sup>nd</sup> stage of the fermentation resulted in higher TAGs accumulation (31%) with no
  by-product accumulation.
  - Overall, results obtained show that Y. lipolytica is a promising biotechnological tool for lipid generation using low-cost acetate media as substrates



Acknowledgments: This work was supported by the European Union's Horizon 2020 research and innovation programme under Grant Agreement No 884208 (BIOfuels production from Syngas FERmentation for Aviation and maritime use - BioSFerA, https://biosfera-project.eu).



H2020-LC-SC3-2018-2019-2020 EUROPEAN COMMISSION Innovation and Networks Executive Agency